



Synthesis and Fluorescence Properties of 4-Cyano and 4-Formyl Melatonin as Putative Melatoninergic Ligands

Silvia Bartolucci, Michele Retini, Fabiola Fanini, Daniele Paderni, and Giovanni Piersanti*

Cite This: ACS Omega 2023, 8, 22190-22194 **Read Online** ACCESS III Metrics & More Article Recommendations Supporting Information ABSTRACT: Fluorescent ligands are imperative to many facets of chemical biology and medicinal chemistry. Herein, we report the syntheses of two fluorescent melatonin-based derivatives as potential ligands of melatonin receptors. The two compounds, namely, 4-cyano and 4-formyl melatonin (4CN-MLT and 4CHO-MLT, respectively), which differ from NHCOCH R = CHO, CN melatonin by only two/three atoms that are very compact in size, were prepared using the selective C3-alkylation of indoles with N-acetyl ethanolamines involving the "borrowing hydrogen" strategy. These compounds exhibit absorption/emission spectra that are red-shifted from MLT

1. INTRODUCTION

Since most biomolecules are either intrinsically nonfluorescent or lack suitable fluorescence properties, one or multiple external fluorophores are frequently required in biological studies using fluorescence-based technology.¹⁻³ Thus, fluorescent probes represent highly sensitive and safe tools for realtime exploration of the activity of biomolecules, visualizing cellular processes, studying ligand/receptor interactions, and more generally increasing the understanding of the pharmacology and physiological processes at the molecular level.⁴⁻¹¹ However, the design and development of such probes are challenging, mainly due to the use of large-sized and interfering fluorophores/organic dyes attached to the ligand (often through a linker), low affinity/specificity of the probe for its target, and extensive nonspecific binding. One practical strategy to alleviate such issues has focused on the use of chromophores that are derived from naturally occurring ligands with a minimally sized reactive group and improved intrinsic optical properties. Considering the rapid progress of fluorescence technologies, effective small-molecule probes containing chromophores have been and still continue to be actively pursued as valuable tools.¹²⁻¹⁴

those of melatonin. Binding studies on two melatonin receptor subtypes showed that these derivatives have a modest affinity and selectivity ratio.

Melatonin (MLT) is an essential human hormone produced mainly by the pineal gland from the amino acid tryptophan; it plays a role in circadian rhythm regulation, acts as an antioxidant, and protects DNA. Despite the importance of MLT in the human body, studies on its interaction with biomolecules, considered explicitly, are quite limited in the literature. In particular, the action of MLT is mediated by two receptors, MT₁ and MT₂, which are members of the huge family of G protein-coupled receptors. These receptors are also implicated in biological processes other than circadian rhythm-and sleep-related disorders,^{15,16} although the potential benefits of MLT assumption in humans remain controversial.¹⁷ Despite the discovery of the high-affinity ligands,¹⁸ research on the pharmacology and the functionality/physiological impact of MLT receptors suffers from the lack of efficient fluorescent probes for these receptors that would allow, among others, studying the molecular dynamics of receptor activation at the molecular level. In addition, given the demanding technical setup, hazards, and expense of using the 2-[125I]-MLT radioligand, fluorescence-based methodologies appear to be more desirable and easier to implement. However, developing such assays for MT_1 and MT_2 depends on the availability of fluorescent tracers, which has been challenging owing to their narrow ligand entry channel and small ligand-binding pocket. The fluorescently labeled melatonin receptor ligands described so far include melatonin-bodipy-fused analogues,¹⁹ 4-azamelatonin ligands with different fluorophores,²⁰ melatonin coupled to the Cy3 cyanin fluorophore,²¹ and coumarin-based compounds,²² but data on their full pharmacological and functional properties are only fragmentary. To offer novel probes, we sought to improve the intrinsic optical properties of the 5-methoxyindole side chain of the endogenous ligand MLT, which possesses fluorescence properties that are

 Received:
 April 13, 2023

 Accepted:
 May 25, 2023

 Published:
 June 8, 2023









unfortunately inappropriate for biological analysis (the main absorption band overlaps with those of the L-tyrosine and L-tryptophan residues already present in proteins), as the source of fluorescence after minor chemical modification, with extended conjugation of the structure that would exhibit absorption at longer wavelengths.^{23–27}

Recent photophysical studies on a library of substituted indoles and tryptophans^{28,29} have demonstrated that the C4 position of the indole ring is special and that the substitutioninduced spectral shift of a substituent may correlate with its electronic withdrawing strength. These reports prompted us to investigate extending the π -conjugation of the indole scaffold at the C4 position of MLT with a small-sized and strong electron-withdrawing group to obtain biologically compatible photophysical properties. Here, we report the design, synthesis, and spectroscopic studies of novel putative fluorescent ligands for MLT receptors that introduce a formyl moiety (CHO) or a cyano (CN) group on the C4 position of the MLT core, which affect the absorption and emission properties compared to the endogenous ligand MLT, without a detrimental effect on binding.^{28,30}

2. RESULTS AND DISCUSSION

Regarding the synthesis of the abovementioned new fluorescent ligands, we envisioned bromide 4 (Scheme 1) as a valuable key intermediate to install both the requisite side chain and the selected substituents in the C4 position of the indole ring. This intermediate was easily obtained from commercially available 5-methoxy-2-carboxyethyl indole (1) by C4-selective bromination (87% yield) followed by saponification and then copper-catalyzed decarboxylation, which allow the preparation of 4 in a good overall yield (65% over two steps). Notably, if the decarboxylation step was performed in the presence of CuCN in dimethylacetamide, direct access to 4-cyano-5-methoxy-1*H*-indole (5) was obtained; otherwise, the same intermediate could also be prepared starting from 4 and using the same reaction.^{31,32}

Pleasingly, the regioselective C3-alkylation of **4** and **5** proceeded uneventfully when *N*-acetyl ethanolamine was employed as the electrophile. It was conducted by taking advantage of the green (water being the only byproduct) borrowing hydrogen process using $[{Cp*IrCl_2}_2]$ as the catalyst (2 mol %) and sodium *tert*-butoxide as the base in 42% yield for **6** and 31% yield for **7**.^{33–36} Suzuki cross-coupling between 7 and potassium vinyl trifluoroborate (vinylBF₃K) was performed to install the vinyl group in good yield (88%).

Oxidative cleavage of the C=C double bond to the corresponding 4CHO-MLT was conducted selectively over the undesired oxidation of the indole C_2-C_3 bond using the Lemieux–Johnson oxidation by employing a catalytic amount of osmium tetroxide and excess sodium periodate in the presence of a non-nucleophilic base, such as 2,6-lutidine. The use of inexpensive commercially available starting materials/reagents is the key feature of this optimized synthetic strategy.

With the synthetic compounds **6** and **9** in hand, we next investigated the optical properties of the synthesized derivatives in different solvents and compared them with those of MLT. A proof of concept of our investigation was gratifyingly obtained directly from the preparation of the target compounds. In fact, blue-light fluorescence could be observed with the naked eye upon illumination of 4CN-MLT with a UV lamp at a wavelength of 254 nm. The UV–Vis spectra of 4CN-MLT and 4CHO-MLT showed absorption bands at 324 and 362 nm, respectively, in aqueous solution (Figure 1).



Figure 1. Absorption spectra of MLT, 4CN-MLT, and 4CHO-MLT, recorded at 10 μ M in the indicated solvents.

Compared to that of MLT, the wavelengths at which 4CN-MLT and 4CHO-MLT had their strongest photon absorption (λ_{max}) showed quite a strong red shift. This finding qualitatively confirmed the expected result that an electron-withdrawing group on an indole ring results in a red shift of λ_{max} . Since the solvent affects the ground and excited state dipoles of a molecule, solvatochromic studies on 4CN-MLT and 4CHO-MLT were conducted in different aprotic and protic polar solvents, such as DMSO, ACN, EtOH, and 1,4-dioxane (Figure 1). No significant shifts in the absorption bands of 6 and 9 were observed in their UV-Vis spectra in these organic solvents. In particular, the λ_{max} of 4CHO-MLT showed a much greater red shift of up to 362 nm in water, whereas that of 4CN-MLT remained constant between 322 and 325 nm and was similar in the different solvents (Table 1).

When 4CN-MLT and 4CHO-MLT were excited at their λ_{max} their fluorescence properties were recorded. It was observed that the emission peaks of both 4CN-MLT and 4CHO-MLT were red shifted compared to that of MLT, as seen for their absorption peaks. Both compounds fluoresce in the visible spectral range, with 4CN-MLT whereas 4CHO-MLT fluoresces in the visible region near 500 nm showing a broad single emission band at around 410 nm (from 396 to 424 nm), whereas 4CHO-MLT. As a result, the absorption spectrum of 4CHO-MLT in ethanol extends beyond 400 nm, making the fluorescence excitable by blue light sources. In addition, the fluorescence spectrum of 4CHO-MLT, which peaks at approximately 480 nm in ethanol with a high intensity, indicates that this MLT derivative is a cyan fluorophore (Figure 2).

For any molecule to be useful as a fluorescent probe for biological spectroscopy and microscopy experiments, other than selective excitation (beyond 360 nm), a strongly red-shifted fluorescence with a large Stokes shift (ca. 120-140 nm), and observable visible fluorescence, it must have a



Figure 2. Emission spectra of MLT, 4CN-MLT, and 4CHO-MLT, recorded at 10 μ M in all studied solvents.

reasonably large fluorescence quantum yield, good photostability, and brightness. Therefore, we also characterized 4CHO-MLT in terms of these properties. As illustrated in the Supporting Information, 4CHO-MLT demonstrated a decent quantum yield (QY = 0.049), a suitable brightness (B = 1120 mM⁻¹ cm⁻¹), and a very good photostability (Figure 3).



Figure 3. Normalized absorption (blue) and fluorescence (red) spectra of 4CHO-MLT (9) in ethanol. The excitation wavelength for the fluorescence measurement was 360 nm. As shown in the inset, 4CHO-MLT (9) is a cyan product.

Finally, it is worth noting that Ladner et al.³⁷ have performed various photoreactions of the indole ring of tryptamine in chloroform to generate new tryptamine-based fluorophores. Interestingly, they found that one of the photoproducts exhibited a green color and a λ_{max} emission of 489 nm. In agreement, the small amount of product obtained corresponded to 9.

Table 1 reports the affinity $[K_i (nM)]$ values of 4CN-MLT and 4CHO-MLT for the human MLT receptors MT₁ and MT₂, which are stably expressed in rat fibroblast NIH3T3 cells, using 2-[¹²⁵I]iodoMLT as the labeled ligand in competition binding assays. Compounds 4CN-MLT and 4CHO-MLT displayed modest affinity for both MLT receptors compared to MLT using a positive control.

Table 1. Affinity $[K_i (nM)]$ Values of 4CN-MLT and 4CHO-MLT for the Human MLT Receptors MT1 and MT2

compound	$MT_1 K_i (nM)$	$MT_2 K_i (nM)$
4CN-MLT	2591 ± 168	398 ± 26
4CHO-MLT	1084 ± 79	424 ± 29
MLT	5.5 ± 0.4	6.3 ± 0.5

As reported recently,^{38,39} the melatonin receptors contain a channel that provides the entrance route of ligands to the binding site from within the lipid bilayer. The atypical (related to metabolite serotonin in 5-HT receptors) entry mechanism could impose constraints on physicochemical properties (i.e., permanent dipole moment) and can be exploited in the future development of novel fluorescent melatonin-based structures to address the need for better binding. Specifically, we imagine that 4-dicyanovinylmelatonin would be a visible chromophore based on the fact that the 2,2-dicyanovinyl group is a stronger electronic withdrawing group than that of nitrile or formyl as well documented in the literature.⁴⁰ In addition, 4CHO-MLT via its aldehydic formyl group could both complement and expand other modes of reactivity (e.g., oxime, hydrazone, and

nitrone formation) to probe unusual and novel fluorescent behavior.

3. CONCLUSIONS

In summary, we have designed, synthesized, biologically evaluated, and surveyed the absorption and emission properties of two four-substituted MLT compounds, namely, 4CN-MLT and 4CHO-MLT, aiming to identify new biologically useful fluorescent melatoninergic ligands. This study provides a convenient means to identify compact MLT derivatives (which are expected to induce minimal perturbation to the native peptide structure, dynamics, or function), whose absorption spectra are red-shifted, and, importantly, whose fluorescence emits visible light. Extension of the π -conjugation of MLT at the C4 position by substitution with a formyl group induced a bathochromic shift to the visible region, and this derivative demonstrated a reasonably large fluorescence quantum yield and photostability. Although these novel compounds revealed only modest binding affinities to the MLT receptors MT1 and MT2, a new and reliable synthetic procedure for the preparation of novel and interesting C4-substituted indoles, which are widely studied by pharmaceutical chemists due to their unique biological activities, was reported.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acsomega.3c02518.

Chemical synthetic methods and ¹H and ¹³C NMR of target compounds and intermediates and spectroscopy (PDF)

AUTHOR INFORMATION

Corresponding Author

Giovanni Piersanti – Department of Biomolecular Sciences, University of Urbino Carlo Bo, 61029 Urbino, Pesaro and Urbino, Italy; o orcid.org/0000-0003-0418-7143; Email: giovanni.piersanti@uniurb.it

Authors

Silvia Bartolucci – Department of Biomolecular Sciences, University of Urbino Carlo Bo, 61029 Urbino, Pesaro and Urbino, Italy; orcid.org/0000-0003-2063-4230

Michele Retini – Department of Biomolecular Sciences, University of Urbino Carlo Bo, 61029 Urbino, Pesaro and Urbino, Italy; • orcid.org/0000-0003-0218-8505

Fabiola Fanini – Department of Biomolecular Sciences, University of Urbino Carlo Bo, 61029 Urbino, Pesaro and Urbino, Italy; orcid.org/0000-0002-8392-2881

Daniele Paderni – Department of Pure and Applied Sciences, University of Urbino Carlo Bo, 61029 Urbino, Pesaro and Urbino, Italy; o orcid.org/0000-0003-4598-5117

Complete contact information is available at: https://pubs.acs.org/10.1021/acsomega.3c02518

Author Contributions

S.B. carried out the study and formatted the manuscript. M.R. assisted in the experiment. F.F. helped to synthesize compounds. M.R., S.B., and D.P. conducted the spectroscopic experiments. G.P. designed the study, analyzed the data, and wrote the manuscript. The manuscript was written through

contributions of all authors. S.B. and M.R. contributed equally to this work.

Funding

This work was supported by the Italian Ministry for University and Research (MUR, PRIN 2020, 2020AEX4TA project) and the University of Urbino grants (DISB_PIERSANTI_PROG_-SIC_ALIMENTARE).

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

We thank the European Union "FSE-REACT-EU, PON Research and innovation 2014–2020 DM 1062/2021" for co-financing the M.R. research contract. We thank Prof. Gilberto Spadoni and Prof. Giorgi Luca for helpful discussions regarding the biological activity and fluorescence properties, respectively. The authors wish to thank Fabrizio Vincenzi and Katia Varani (Department of Translational Medicine, University of Ferrara, Via Luigi Borsari 46, 44121, Ferrara, Italy) for the binding affinity studies on MLT receptors.

REFERENCES

(1) For reviews, see: (a) Chinen, A. B.; Guan, C. M.; Ferrer, J. R.; Barnaby, S. N.; Merkel, T. J.; Mirkin, C. A. Nanoparticle Probes for the Detection of Cancer Biomarkers, Cells and Tissues by Fluorescence. *Chem. Rev.* **2015**, *115*, 10530–10574.

(2) Chen, X.; Wang, F.; Hyun, J. Y.; Wei, T.; Qiang, J.; Ren, X.; Shin, I.; Yoon, J. Recent Progress in the Development of Fluorescent, Luminescent and Colorimetric Probes for Detection of Reactive Oxygen and Nitrogen Species. *Chem. Soc. Rev.* **2016**, *45*, 2976–3016.

(3) Benson, S.; de Moliner, F.; Tipping, W.; Vendrell, M. Miniaturized Chemical Tags for Optical Imaging. *Angew. Chem., Int. Ed.* **2022**, *61*, No. e202204788.

(4) Kuder, K.; Kieć-Kononowicz, K. Fluorescent GPCR Ligands as New Tools in Pharmacology. *Curr. Med. Chem.* 2008, *15*, 2132–2143.
(5) Böhme, I.; Beck-Sickinger, A. G. Illuminating the Life of GPCRs.

Cell Commun. Signal. 2009, 7, 16.

(6) Goddard, A. D.; Watts, A. Contributions of Fluorescence Techniques to Understanding G Protein-Coupled Receptor Dimerisation. *Biophys. Rev.* 2012, 4, 291–298.

(7) Jakobs, D.; Sorkalla, T.; Häberlein, H. Ligands for Fluorescence Correlation Spectroscopy on g Protein-Coupled Receptors. *Curr. Med. Chem.* **2012**, *19*, 4722–4730.

(8) Cottet, M.; Faklaris, O.; Zwier, J. M.; Trinquet, E.; Pin, J.-P.; Durroux, T. Original fluorescent ligand-based assays open new perspectives in G-protein coupled receptor drug screening. *Pharmaceuticals* **2011**, *4*, 202–214.

(9) Ma, Z.; Du, L.; Li, M. Toward Fluorescent Probes for G-Protein-Coupled Receptors (GPCRs). J. Med. Chem. 2014, 57, 8187–8203.

(10) Borgarelli, C.; Klingl, Y. E.; Escamilla-Ayala, A.; Munck, S.; Van Den Bosch, L.; De Borggraeve, W. M.; Ismalaj, E. Lighting Up the Plasma Membrane: Development and Applications of Fluorescent Ligands for Transmembrane Proteins. *Chem. – Eur. J.* **2021**, *27*, 8605–8641.

(11) Suofu, Y.; Li, W.; Jean-Alphonse, F. G.; Jia, J.; Khattar, N. K.; Li, J.; Baranov, S. V.; Leronni, D.; Mihalik, A. C.; He, Y.; Cecon, E.; Wehbi, V. L.; Kim, J. H.; Heath, B. E.; Baranova, O. V.; Wang, X.; Gable, M. J.; Kretz, E. S.; Di Benedetto, G.; Lezon, T. R.; Ferrando, L. M.; Larkin, T. M.; Sullivan, M.; Yablonska, S.; Wang, J.; Minnigh, M. B.; Guillaumet, G.; Suzenet, F.; Richardson, R. M.; Poloyac, S. M.; Stolz, D. B.; Jockers, R.; Witt-Enderby, P. A.; Carlisle, D. L.; Vilardaga, J. P.; Friedlander, R. M. Dual Role of Mitochondria in Producing Melatonin and Driving GPCR Signaling to Block Cytochrome c Release. *Proc. Natl. Acad. Sci. U. S. A.* **2017**, *114*, E7997–E8006. (12) Sinkeldam, R. W.; Greco, N. J.; Tor, Y. Fluorescent Analogs of Biomolecular Building Blocks: Design Properties and Applications. *Chem. Rev.* **2010**, *110*, 2579–2619.

(13) Niu, L.-Y.; Chen, Y.-Z.; Zheng, H.-R.; Wu, L.-Z.; Tung, C.-H.; Yang, Q.-Z. Design Strategies of Fluorescent Probes for Selective Detection Among Biothiols. *Chem. Soc. Rev.* **2015**, *44*, 6143–6160.

(14) Singh, H.; Tiwari, K.; Tiwari, R.; Pramanik, S. K.; Das, A. Small Molecule as Fluorescent Probes for Monitoring Intracellular Enzymatic Transformations. *Chem. Rev.* **2019**, *119*, 11718–11760.

(15) Spadoni, G.; Bedini, A.; Rivara, S.; Mor, M. Melatonin Receptor Agonists: New Options for Insomnia and Depression Treatment. *CNS Neurosci. Ther.* **2011**, *17*, 733–741.

(16) Delagrange, P.; Atkinson, J.; Boutin, J. A.; Casteilla, L.; Lesieur, D.; Misslin, R.; Pellissier, S.; Pénicaud, L.; Renard, P. Therapeutic Perspectives for Melatonin Agonists and Antagonists. *J. Neuro-endocrinol.* **2003**, *15*, 442–448.

(17) Boutin, J. A.; Jockers, R. Melatonin controversies, an update. J. Pineal Res. 2021, 70, No. e12702.

(18) Boutin, J. A.; Witt-Enderby, P. A.; Sotriffer, C.; Zotlos, D. P. Melatonin receptor ligands: A pharmaco-chemical perspective. *J. Pineal Res.* **2020**, *69*, No. e12672.

(19) Thireau, J.; Marteaux, J.; Delagrange, P.; Lefoulon, F.; Dufourny, L.; Guillaumet, G.; Suzenet, F. Original Design of Fluorescent Ligands by Fusing BODIPY and Melatonin Neuro-hormone. *ACS Med. Chem. Lett.* **2014**, *5*, 158–161.

(20) Viault, G.; Poupart, S.; Mourlevat, S.; Lagaraine, C.; Devavry, S.; Lefoulon, F.; Bozon, V.; Dufourny, L.; Delagrange, P.; Guillaumet, G.; Suzenet, F. Design, Synthesis and Biological Evaluation of Fluorescent Ligands for MT_1 and/or MT_2 Melatonin Receptors. *RSC Adv.* **2016**, *6*, 62508–62521.

(21) Gbahou, F.; Cecon, E.; Viault, G.; Gerbier, R.; Jean-Alphonse, F.; Karamitri, A.; Guillaumet, G.; Delagrange, P.; Friedlander, R. M.; Vilardaga, J. P.; Suzenet, F.; Jockers, R. Design and Validation of the First Cell-Impermeant Melatonin Receptor Agonist. *Br. J. Pharmacol.* **2017**, *174*, 2409–2421.

(22) De la Fuente Revenga, M.; Herrera-Arozamena, C.; Fernández-Sáez, N.; Barco, G.; García-Orue, I.; Sugden, D.; Rivara, S.; Rodríguez-Franco, M. I. New coumarin-based fluorescent melatonin ligands. Design, synthesis and pharmacological characterization. *Eur. J. Med. Chem.* **2015**, *103*, 370–373.

(23) Bartoccini, F.; Bartolucci, S.; Mari, M.; Piersanti, G. A Simple, Modular Synthesis of C4-Substituted Tryptophan Derivatives. *Org. Biomol. Chem.* **2016**, *14*, 10095–10100.

(24) Boville, C. E.; Romney, D. K.; Almhjell, P. J.; Sieben, M.; Arnold, F. H. Improved Synthesis of 4-Cyanotryptophan and other Tryptophan Analogues in Aqueous Solvents Using Variants of TrpB from Thermotoga maritima. *J. Org. Chem.* **2018**, *83*, 7447–7452.

(25) Talukder, P.; Chen, S.; Roy, B.; Yakovchuk, P.; Spiering, M. M.; Alam, M. P.; Madathil, M. M.; Bhattacharya, C.; Benkovic, S. J.; Hecht, S. M. Cyanotryptophans as Novel Fluorescent Probes for Studying Protein Conformational Changes and DNA-Protein Interaction. *Biochemistry* **2015**, *54*, 7457–7469.

(26) Hilaire, M. R.; Ahmed, I. A.; Lin, C.-W.; Jo, H.; DeGrado, W. F.; Gai, F. Blue Fluorescent Amino Acid for Biological Spectroscopy and Microscopy. *Proc. Natl. Acad. Sci. U. S. A.* **2017**, *114*, 6005–6009. (27) Zhang, K.; Ahmed, I. A.; Kratochvil, H. T.; DeGrado, W. F.; Gai, F.; Jo, H. Synthesis and Application of the Blue Fluorescent Amino Acid L-4-Cyanotryptophan to Assess Peptide-Membrane Interactions. *Chem. Commun.* **2019**, *55*, 5095–5098.

(28) Micikas, R. J.; Ahmed, I. A.; Acharyya, A.; Smith, A. B., III; Gai, F. Tuning the electronic transition energy of indole via substitution: application to identify tryptophan-based chromophores that absorb and emit visible light. *Phys. Chem. Chem. Phys.* **2021**, *23*, 6433–6437.

(29) Acharyya, A.; Zhang, W.; Gai, F. Tryptophan as a Template for Development of Visible Fluorescent Amino Acids. J. Phys. Chem. B 2021, 125, 5458–5465.

(30) A-Value (kcal/mol) 0.17 and 0.8; sigma value around 0.6 and 0.4; Hansch, C.; Leo, A.; Taft, R. W. A Survey of Hammett

Substituent Constants and Resonance and Field Parameters. *Chem. Rev.* **1991**, *91*, 165–195.

(31) Meyer, M. D.; Kruse, L. I. Ergoline synthons: Synthesis of 3,4dihydro-6-methoxybenz[cd]indol-5(1H)-one (6-methoxy-Uhle's ketone) and 3,4-dihydrobenz[cd]indol-5(1H)-one (Uhle's ketone) via a novel decarboxylation of indole-2-carboxylates. *J. Org. Chem.* **1984**, *49*, 3195–3199.

(32) Kruse, L. I.; Meyer, M. D. Ergoline synthons. 2. Synthesis of 1,5-dihydrobenz[cd]indol-4(3H)-ones and 1,3,4,5-tetrahydrobenz-[cd]indol-4-amines. J. Org. Chem. **1984**, 49, 4761–4768.

(33) Bartolucci, S.; Mari, M.; Bedini, A.; Piersanti, G.; Spadoni, G. Iridium-Catalyzed Direct Synthesis of Tryptamine Derivatives from Indoles: Exploiting N-Protected β -Amino Alcohols as Alkylating Agents. J. Org. Chem. **2015**, 80, 3217–3222.

(34) Bartolucci, S.; Mari, M.; Di Gregorio, G.; Piersanti, G. Observations concerning the synthesis of tryptamine homologues and branched tryptamine derivatives via the borrowing hydrogen process: synthesis of psilocin, bufotenin, and serotonin. *Tetrahedron* **2016**, *72*, 2233–2238.

(35) Bartoccini, F.; Retini, M.; Piersanti, G. C3-Alkylation of indoles and oxindoles by alcohols by means of borrowing hydrogen methodology. *Tetrahedron Lett.* **2020**, *61*, No. 151875.

(36) Hall, C. J. J.; Goundry, W. R. F.; Donohoe, T. J. Hydrogen-Borrowing Alkylation of 1,2-Amino Alcohols in the Synthesis of Enantioenriched γ -Aminobutyric Acids. *Angew. Chem., Int. Ed.* **2021**, *60*, 6981–6985.

(37) Ladner, C. L.; Tran, K.; Le, M.; Turner, R. J.; Edwards, R. A. Excited state photoreaction between the indole side chain of tryptophan and halocompounds generates new fluorophores and unique modifications. *Photochem. Photobiol.* **2014**, *90*, 1027–1033.

(38) Stauch, B.; Johansson, L. C.; McCorvy, J. D.; Patel, N.; Han, G. W.; Huang, X.; Gati, C.; Batyuk, A.; Slocum, S. T.; Ishchenko, A.; Brehm, W.; White, T. A.; Michaelian, N.; Madsen, C.; Zhu, L.; Grant, T. D.; Grandner, J. M.; Shiriaeva, A.; Olsen, R. H. J.; Tribo, A. R.; Yous, S.; Stevens, R. C.; Weierstall, U.; Katritch, V.; Roth, B. L.; Liu, W.; Cherezov, V. Structural basis of ligand recognition at the human MT₁ melatonin receptor. *Nature* **2019**, *569*, 284–288.

(39) Johansson, L. C.; Stauch, B.; McCorvy, J. D.; Han, G. W.; Patel, N.; Huang, X.; Batyuk, A.; Gati, C.; Slocum, S. T.; Li, C.; Grandner, J. M.; Hao, S.; Olsen, R. H. J.; Tribo, A. R.; Zaare, S.; Zhu, L.; Zatsepin, N. A.; Weierstall, U.; Yous, S.; Stevens, R. C.; Liu, W.; Roth, B. L.; Katritch, V.; Cherezov, V. XFEL structures of the human MT₂ melatonin receptor reveal the basis of subtype selectivity. *Nature* **2019**, 569, 289–292.

(40) Micikas, R. J.; Acharyya, A.; Smith, A.B., III; Gai, F. Synthesis and characterization of the fluorescence utility of two visible-lightabsorbing tryptophan derivatives. *Chem. Phys. Lett.* **2022**, 795, No. 139553. and references cited therein.