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3 1 **Nutritional, antioxidant and sensory properties of functional beef burgers formulated with chia**  
4 **seeds and goji puree, before and after *in vitro* digestion**  
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8 4 Elena Antonini<sup>a\*</sup>, Luisa Torri<sup>b</sup>, Maria Piochi<sup>b</sup>, Giorgia Cabrino<sup>b</sup>, Maria Assunta Meli<sup>a</sup>, Roberta De  
9 Bellis<sup>a</sup>  
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13 7 <sup>a</sup>Department of Biomolecular Sciences, University of Urbino Carlo Bo, via Saffi 2, 61029 Urbino (PU),  
14 Italy  
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16 9 <sup>b</sup>University of Gastronomic Sciences, Piazza Vittorio Emanuele 9, 12060 Bra (CN), Italy  
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19 11 **\*Corresponding author**

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21 12 e-mail address: elena.antonini@uniurb.it (E. Antonini)  
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62 **Abstract**  
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64 14 The addition of chia seeds and goji puree (2.5 and/or 5%) was evaluated in terms of their effects on the  
65 15 fatty acid profile, lipid peroxidation, total phenols and antioxidant capacity of cooked beef burgers. In  
66 16 comparison to control burgers, polyunsaturated fatty acids doubled or tripled in samples containing  
67 17 chia seeds; polyphenols and antioxidant capacities (ORAC, ABTS, DPPH) increased up to 70% and  
68 18 malondialdehyde values were reduced up to 50% in burgers formulated with both ingredients.

69 19 Polyphenols, antioxidant capacity and lipid peroxidation were also assessed after *in vitro* digestion. A  
70 20 marked increase of polyphenol bioaccessibility and antioxidant capacity was observed for all samples,  
71 21 but also malondialdehyde values were increased after digestion, especially in samples containing 5%  
72 22 chia seeds.

73 23 Finally, hedonistic tests were conducted on young (18-30 years), adult (31-60 years) and elderly (>60  
74 24 years) subjects and the burgers resulted acceptable by all groups, appointing to their potential  
75 25 application as functional burgers.  
76 26

77 27 **Keywords:** Functional beef burgers; chia seeds; goji puree; natural antioxidants; lipid peroxidation; *in*  
78 28 *vitro* digestion.  
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## 1. Introduction

Meat represents a good source of proteins with high biological value and various micronutrients, including vitamin B12, zinc, phosphorus and iron (De Smet & Vossen, 2016). Nevertheless, in the past two decades, the consumption of red meat and meat products has been stigmatized due to their saturated fat, cholesterol and salt contents, which are considered as the main risk factors for several chronic diseases (De Smet & Vossen, 2016).

Despite this, the most recent findings suggest that dietary policies should encourage the intake of vegetables or foods rich in polyunsaturated fatty acids, whose consumption is lower than the optimal levels, more than reducing the consumption of red meat or processed meat. This might have a greater health effect than dietary policies only targeting sugar and fat (Afshin et al., 2019).

In Italy, the average annual consumption of beef meat has fallen from 24.9 (2007) to 17.1 (2017) kg per capita. However, in 2018, an increase in red meat consumption (+5%) was registered, with consumers paying more attention to the quality, genuineness and possible health benefits of meat (ISMEA, 2018).

Several strategies have been proposed to improve the quality of meat, not only at the level of breeding, but also during processing, to develop functional or nutraceutical meat products and provide adequate responses to consumer requirements (Decker & Park, 2010; Olmedilla-Alonso, Jimenez-Colmenero, & Sanchez-Muniz, 2013). Such strategies include the addition of natural antioxidants, able to prevent lipid peroxidation, which causes the deterioration of meat quality (Falowo, Fayemi, & Muchenje, 2014; Kumar, Yadav, Ahmad, & Narsaiah, 2015; Ribeiro et al., 2019) or vegetable oils or oilseeds, able to improve the fatty acid profile of meat (Decker & Park, 2010). Because of their promising functional properties, chia seeds and goji berries have gained attention as potential healthy ingredients for developing new functional foods.

The popularity of chia seeds (*Salvia hispanica* L.) has grown rapidly in the last few years because of their health-promoting activities, which include cardio-protective, antioxidant, anticancer and antimicrobial effects (Muñoz, Cobos, Diaz, & Aguilera, 2013; Ullah et al., 2016). In particular, chia seeds have high lipid, polyphenol and fiber contents and represent the source with the highest concentration of  $\omega$ -3 alpha-linolenic acid (ALA) and  $\omega$ -6 linoleic acid (Ayerza & Coates, 2011).

Goji berries (*Lycium barbarum* and *Lycium chinense* fruits) are commonly consumed in their dried form. Recently, they have become increasingly popular as “superfruits” because of their potential health-promoting properties, including antioxidant, hypoglycemic, lipid-lowering, immunostimulatory and anticancer effects (Chang, Alasalvar, & Shahidi, 2018; Kulczyński & Gramza-Michałowska, 2016; Ma et al., 2019). The main bioactive compounds in goji berries are water-soluble and highly branched polysaccharides; carotenoids, which confer the orange-red color to the berries; and phenolic

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180 62 compounds and flavonoids, which have a very high antioxidant capacity (Kulczyński & Gramza-  
181 63 Michałowska, 2016).

183 64 Although goji berries and chia seeds have been widely recognized as potentially healthy **foods**, few  
184 64 studies have reported their application in meat products (Aco, Aco, & Elena, 2018; Bulambaeva,  
185 65 Vlahova-Vangelova, Dragoev, Balev, & Uzakov, 2014; Pintado, Herrero, Jiménez-Colmenero,  
186 66 Pasqualin Cavalheiro, & Ruiz-Capillas, 2018; Pintado, Herrero, Jiménez-Colmenero, & Ruiz-Capillas,  
187 67 2016; Souza et al., 2015; Zaki, 2018). To the best of our knowledge, no studies have investigated the  
188 68 combination of goji berries, in a puree form, and chia seeds, used as sources of natural antioxidants and  
189 69 polyunsaturated fatty acids to formulate beef burgers. Hence, the aim of the present study was to the  
190 70 evaluate the effects of the addition of chia seeds and/or goji puree, used alone or in combination at 2.5  
191 71 and 5%, on the physicochemical and nutritional properties of cooked beef burgers. An overall  
192 72 screening of the antioxidant properties of the realized burgers was also performed by measuring their  
193 73 total phenol content, antioxidant capacities (ORAC, ABTS and DPPH assays) and malondialdehyde  
194 74 (MDA, a secondary product of lipid peroxidation) levels.

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202 76 In order to be able to confer health benefits, bioactive compounds must resist food processing and be  
203 77 bioavailable, i.e. be released from the food matrix and be bioaccessible in the gastrointestinal tract  
204 78 (Angelino et al., 2017; Espín, García-Conesa, & Tomás-Barberán, 2007; Rein et al., 2013). We  
205 79 therefore applied an *in vitro* digestion model, characterized by an oral, gastric and intestinal phase, to  
206 80 evaluate the bioaccessibility of antioxidants and their ability to counteract lipid peroxidation in digested  
207 81 cooked burgers.

208 82 Finally, as the addition of ingredients potentially able to improve the nutritional quality of products  
209 83 may also alter their sensory properties, a hedonic test of our products was performed among different  
210 84 groups of consumers (young, adults and elderly subjects), in order to evaluate the overall liking of the  
211 85 burgers.

## 219 86 **2. Materials and methods**

### 220 87 **2.1. Materials**

221 88 Beef and vegetable dietary fibers (fibers 84 %; botanical origin: carrot, bamboo, potato, *Plantago spp.*)  
222 89 were supplied by the Company Baldi srl (Jesi, AN, Italy). Goji puree, made from fresh goji berries  
223 90 grown in Italy, was supplied by Rete di Imprese “Likion” per la Filiera del Goji Italiano®, (Villa San  
224 91 Giovanni, RC, Italy). Chia seeds were purchased from a local market. The average nutritional values of  
225 92 the goji puree and chia seeds, along with their moisture and pH values are reported in Table S1.

226 93 AAPH (2,2'-azobis(2-methylpropionamide) dihydrochloride),  $\alpha$ -amylase, BHT (2,6-di-tert-butyl-4-  
227 94 methylphenol), bile, BSA (bovine serum albumin), DPPH (2,2-diphenyl-1-picrylhydrazyl), fluorescein

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298 127 as the percentage of total fatty acids determined. The atherogenicity (AI) and thrombogenicity (TI)  
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300 128 indexes were calculated as reported in the literature (Ulbricht & Southgate, 1991).

## 301 302 129 **2.5. In vitro digestion**

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304 130 The *in vitro* digestion (oral, gastric and intestinal phases) was performed on cooked burgers, in  
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306 131 triplicate, following the procedure reported in the literature (Ninfali, Mari, Meli, Roselli, & Antonini,  
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308 132 2019) with slight modifications (Table S2). The final suspension derived from the intestinal phase ( $\approx$   
309 133 35 mL) was used “as-is” for the malondialdehyde (MDA) measurement or centrifuged at 15,000 rpm at  
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311 134 +4 °C for 30 min and filtered using 0.45  $\mu$ m filters for the polyphenol, ORAC, ABTS and DPPH  
312 135 assays. A blank sample was prepared with the digestive juices only.

## 313 314 136 **2.6. Extraction and determinations of total phenols (TPs)**

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316 137 The extraction of TPs from goji puree, pulverized chia seeds and cooked burgers was performed in  
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318 138 triplicate, as previously reported by Wu, Duckett, Neel, Fontenot, and Clapham (2008). TPs were  
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320 139 assayed using the Folin-Ciocalteu method, as previously reported by Singleton, Orthofer, and Lamuela-  
321 140 Raventós (1999). A calibration curve was prepared using gallic acid (from 2 to 10  $\mu$ g/mL) as a standard  
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323 141 and values were expressed as mg of gallic acid equivalents (GAE)/100 g product.

## 324 142 **2.7. Antioxidant assays (ORAC, DPPH, ABTS)**

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326 143 Three assays were performed to examine the antioxidant capacity of TPs in all samples: the oxygen  
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328 144 radical absorbance capacity (ORAC), the DPPH and the ABTS radical scavenging activity assays. Each  
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330 145 assay was performed in triplicate.

331 146 The ORAC method was performed as previously reported by Ninfali et al. (2009). The DPPH and  
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333 147 ABTS assays were performed as reported by Brand-Williams, Cuvelier, and Berset (1995) and Ferri,  
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335 148 Gianotti, and Tassoni (2013), respectively. A calibration curve was prepared for each assay (ORAC,  
336 149 DPPH, ABTS) with Trolox (from 2 to 20  $\mu$ M) used as a standard and values expressed as  $\mu$ mol of  
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338 150 Trolox equivalents (TE)/100 g product.

## 339 151 **2.8. Measurement of malondialdehyde (MDA)**

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342 152 The extraction and quantification of malondialdehyde (MDA) in cooked burgers, before and after  
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344 153 digestion, was performed in triplicate, following the procedure reported by Jung, Nam, and Jo (2016),  
345 154 with slight modifications. Briefly, 4.5 g of digested and non-digested cooked burgers were  
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347 155 homogenized with 30 ml of 7.5% TCA solution and 150  $\mu$ L of 7.2% BHT in ethanol using an Ultra-  
348 156 Turrax<sup>®</sup> at 16,000 rpm for 1 min. The homogenate was centrifuged at 10,000 rpm for 15 min, filtered  
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350 157 with 20-25  $\mu$ m filters and used as the MDA extract. For its quantification, 1 mL of MDA extract was  
351 158 mixed with 1 mL of 20 mM TBA in screw-cap tubes. The tubes were heated in a boiling water bath at  
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159 90 °C for 30 min and cooled in ice. Absorbance was measured at 532 nm using the UV–vis  
160 spectrophotometer. A calibration curve was prepared using TEP (1-32 µM) and results were expressed  
161 as mg of MDA/100 g product.

## 2.9. Sensory evaluation

163 Sensory tests were conducted on three different groups of subjects: young (18-30 years; n=56;  
164 M=48%), adult (31-60 years; n=38, M=74%) and elderly (>60 years; n=33, M=39%). The young  
165 subjects were recruited at the University of Gastronomic Sciences (Bra, CN, Italy), the adults were  
166 recruited at the Baldi srl Company and the elderly subjects joined the test at the retirement home “Casa  
167 di riposo – Residenza protetta Hermes di proprietà della Fondazione Opere Laiche Lauretane e Casa  
168 Hermes” (Loreto, AN, Italy). The sensory study was approved by the Ethics Committee of the  
169 University of Gastronomic Sciences. Written informed consent was collected from participants prior to  
170 the test.

171 Four samples (CTRL, G<sub>2.5</sub>, C<sub>2.5</sub>, G<sub>2.5</sub>+C<sub>2.5</sub>) were evaluated. Cooked burgers were divided into  
172 approximately 20 g portion and served in transparent plastic containers sealed with a lid. Samples were  
173 codified with three-digit random codes for young and adult subjects and with two-digit random codes  
174 for elderly subjects. The serving order was randomized and balanced across assessors.

175 Subjects were required to observe, smell and taste all the samples and give a judgment of liking  
176 considering appearance, odor, taste, flavor, texture and overall liking. Liking was evaluated on a nine-  
177 point hedonic scale ranging from 1 = extremely dislike to 9 = extremely like (Peryam & Pilgrim, 1957).  
178 A 30-sec break was required among samples combined with a rinsing procedure with water. Plain  
179 bread was given to the elderly subjects, while non-salted crackers were given to the young and adult  
180 groups as palate cleansers.

181 Participants were asked to complete a short questionnaire, which included questions regarding the  
182 frequency of consumption of red/white meat, consumption of cured meats, preferred type of meat (red,  
183 white or cured meat). Considering the frequency of meat consumption, subjects were classified in three  
184 clusters: occasional (once a week or less), frequent (2-3 times per week) and very frequent (at least 4  
185 times per week). The tests lasted approximately from 10 min (young) to over 40 min (elderly). Data  
186 were collected with an automated procedure (FIZZ Acquisition software, version 2.51C, Biosystèmes,  
187 Courtenon, France) for the youngest, while paper sheets were used for the adults and elderly subjects.

## 2.10. Statistical analysis

189 The statistical significance ( $P < 0.05$ ) of the effect of sample formulation was tested by one-way  
190 analysis of variance (ANOVA) using the SPSS® 17.0 statistical package program (IBM, Chicago, IL,  
191 USA). The entire experiment was performed in triplicate on three different days and no statistically

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416 192 significant differences were found between replicates. Differences between means for formulations  
417 193 were compared using Bonferroni's test.

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420 194 The effect of the subject group (young, adult, elderly) on liking for the four prototypes was assessed  
421 195 adopting two-way mixed ANOVA models (fixed factor: sample; random factor: group) separately for  
422 196 appearance, odor, taste, flavor, texture and overall liking. Two-ANOVA mixed models (fixed factor:  
423 197 sample; random factor: assessor) were separately applied within each group and on the whole  
424 198 population to assess the effect of the sample on liking for each sensory modality, followed by Fisher's  
427 199 LSD test to estimate significant differences among mean values ( $P < 0.05$ ). The effect of the frequency  
429 200 of consumption on the liking of samples was assessed with two-way mixed ANOVA models (fixed  
430 201 factor: frequency of consumption; random factor: assessor). Sensory analyses were conducted with  
432 202 XLStat 2019.1.1, Addinsoft, Boston, USA.

### 434 203 **3. Results and discussion**

#### 435 204 **3.1. Proximate composition, pH and cooking loss**

437  
438 205 Table 2 shows the proximate composition, pH and cooking loss (CL) of burgers made with different  
439 206 percentages of goji puree and chia seeds, used alone or in combination.

441 207 The moisture content ranged from 64.7 to 60.7% (Table 2). The addition of 5% chia seeds significantly  
442 208 decreased the moisture content of C<sub>5</sub> and G<sub>5</sub>+C<sub>5</sub> burgers due to the increase of dry matter in the  
444 209 formulations. In fact, the chia seeds used in our experiments were characterized by only 5.5% moisture,  
445 210 a significantly lower value than that of goji puree (76.8%) (Table S1).

447 211 The fat content ranged from 11.8 to 14.7% (Table 2), with the highest level found in C<sub>5</sub> burgers due to  
448 212 the presence of chia seeds, which were characterized by a fat content of 35% (Table S1). The addition  
450 213 of up to 5% goji puree and chia seeds did not influence the protein content, which showed an average  
452 214 value of 22.8% with no significant differences among samples (Table 2).

453 215 The pH values ranged from 5.9 to 5.7 (Table 2). The addition of 5% goji puree led to a greater decrease  
455 216 in the pH values of G<sub>5</sub> and G<sub>5</sub>+C<sub>5</sub> burgers due to its acidic nature (pH = 4.8) compared to chia seeds  
456 217 (Table S1).

458 218 CL varied from 26.1 to 16.7% (Table 2). Burgers containing chia seeds showed smaller losses than the  
459 219 other samples due to the high water retention and emulsifying capacities of the mucilaginous  
460 220 compounds produced by glucuronic acid and neutral sugars, which constitute the soluble fiber of chia  
463 221 seeds (de Melo et al., 2015).

#### 465 222 **3.2. Fatty acid composition**

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467 223 Table 3 shows the fatty acid composition of cooked burgers and their nutritionally significant ratios. In  
468 224 all samples, the main fatty acids were monounsaturated (MUFA), followed by saturated (SFA) and  
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polyunsaturated (PUFA). Oleic acid (C18:1) predominated among MUFA, whereas palmitic acid (C16:0) was the main acid among SFA.

The addition of chia seeds enhanced the nutritional quality of the beef burgers (Table 3), resulting in:

- i) an increase in total PUFA content, which ranged from 2.9% (CTRL) to 6.9% (mean value of C<sub>2.5</sub> and G<sub>2.5</sub>+C<sub>2.5</sub>) and 10.5% (mean value of C<sub>5</sub> and G<sub>5</sub>+C<sub>5</sub>), due to the high levels of  $\alpha$ -linolenic acid (C18:3) present in the chia seeds;
- ii) an improved PUFA/SFA ratio, which increased from 0.06 (CTRL) to 0.26 (G<sub>5</sub>+C<sub>5</sub>). Considering the medium-high fat content of the beef used to formulate the burgers, compared to studies in the literature, the improved PUFA/SFA ratio should be considered of interest, even though it is half the recommended ratio of 0.4 (Wood et al., 2004);
- iii) an improved  $\omega$ -6/ $\omega$ -3 ratio, decreasing from 5.67 (CTRL) to 0.65 (G<sub>5</sub>+C<sub>5</sub>). This ratio plays a very important role in human nutrition, more so than the ratio reported in the previous point. According to nutritional guidelines, the  $\omega$ -6/ $\omega$ -3 ratio should not be higher than 4 (Simopoulos, 2002);
- iv) a reduction of atherogenicity (AI) and thrombogenicity (TI) indexes, ranging from 0.60 and 1.56 (CTRL) to 0.51 and 0.87 (C<sub>5</sub>), respectively. These indexes take into account the different effects of fatty acids on cardiovascular risk: food products with low values of AI and TI can inhibit the aggregation of platelets and decrease the levels of esterified fatty acids, cholesterol and phospholipids, thereby lowering the risk of micro- and macro-coronary disease (Ulbricht & Southgate, 1991).

The improved lipid profile obtained by adding chia seeds to beef burgers can play an important role in the prevention and treatment of cardiovascular diseases, hypertension, diabetes, arthritis and other autoimmune disorders (Cifuni, Napolitano, Riviezzi, Braghieri, & Girolami, 2004).

In addition, the fatty acid profile of our burgers formulated with chia seeds made it possible to satisfy EU Regulation n° 116/2010 regarding the claims “source of omega-3 fatty acids” and “high omega-3 fatty acids”, which can be used if the product contains at least 0.3 g or 0.6 g  $\alpha$ -linolenic acid (ALA) per 100 g, respectively. According to our results, C<sub>2.5</sub> and G<sub>2.5</sub>+C<sub>2.5</sub> contain an average amount of 0.49 g ALA/100 g; C<sub>5</sub> and G<sub>5</sub>+C<sub>5</sub> contain about 0.91 g ALA/100 g. This fatty acid contributes to the maintenance of normal blood cholesterol levels and this beneficial effect is obtained with a daily intake of 2 g of ALA (EU Reg. 432/2012).

### **3.3. Polyphenol contents and antioxidant profiles of cooked beef burgers before and after *in vitro* digestion**



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In this study, we first evaluated the antioxidant profiles of the goji puree and chia seeds used in the formulation of beef burgers (Table 4). Both plant-based ingredients were characterized by similar total phenols, which could be ascribed to their pool of antioxidant molecules belonging to the same family, i.e. carotenoids, chlorogenic and caffeic acids, quercetin, and kaempferol (Kulczyński & Gramza-Michałowska, 2016; Ullah et al., 2016). Nevertheless, the food matrix of each product influenced its antioxidant capacity in different ways, with the goji berry puree showing higher ORAC and ABTS values and the chia seeds showing higher DPPH values (Table 4).

Moreover, meat itself contains hydrophilic and lipophilic antioxidants, which moderately contribute to its antioxidant capacity. Among these, the most efficient antioxidant compounds are dipeptides such as carnosine and anserine, and other substances such as L-carnitine, glutathione, taurine and creatine (Liu, Xing, Fu, Zhou, & Zhang, 2016; M Antonini et al., 2002). Before digestion, the polyphenol content of our CTRL burger was 20.9 mg/100g (Fig. 1A, blank bar). This value increased to 25 and 29 mg/100g when the goji puree and chia seeds were added to the formulation at 2.5% (G<sub>2.5</sub> and C<sub>2.5</sub>) and 5% (G<sub>5</sub> and C<sub>5</sub>), respectively. The combination of the two ingredients at 5% (G<sub>5</sub>+C<sub>5</sub>) showed the highest polyphenol level (34 mg/100 g), thus suggesting that the antioxidant molecules that characterize goji berries and chia seeds have a synergistic effect (Fig. 1A, blank bars). To the best of our knowledge, in literature there are not studies regarding the possible synergistic effect of these two healthy foods, although interactions between phytochemicals from fruits and vegetables have been recently reviewed (Phan, Paterson, Bucknall, & Arcot, 2018). The two main methods used to determine the types of interaction of binary mixtures of phytochemicals, i.e. isobologram and combination index, reported in this review, could be used in future studies to test the possible interaction between phytochemicals of chia seeds and goji puree.

Polyphenol bioaccessibility is an important parameter that represents the amount of polyphenols which are released from the food matrix, strongly influenced by the physicochemical properties of the food matrix and by the technological processes used in food production, as well as by the physiological condition of the individual (Angelino et al., 2017). After digestion, the polyphenol content showed a ten-fold increase in most of the analyzed burgers (Fig 1A, grey bars) due to the further solubilization of polyphenols in digestive fluids (Kim & Hur, 2018; Pešić et al., 2019). Digestive enzymes are able to transform phenolic compounds into different structural forms possessing altered chemical properties. The increase in the total phenolic content may be attributed to an acidic hydrolysis of phenolic glycosides during gastric digestion, with a higher antioxidant activity displayed by aglycone phenolics than their glycoside forms (Lee, Lee, Chung, & Hur, 2016). Moreover, the change of pH from an acidic to the alkaline environment, during intestinal digestion, may improve the antioxidant capacity of

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593 290 phenolics through the deprotonation of the hydroxyl moieties present on their aromatic ring (Kim &  
594 291 Hur, 2018).

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596 292 Nevertheless, the polyphenol pattern of digested burgers (Fig 1A, grey bars) was not the same as that of  
597 292 non-digested burgers (Fig 1A, blank bars). The best results were found in G<sub>2.5</sub> and G<sub>5</sub> digested burgers  
598 293 (average value of 245 mg/100g), while C<sub>5</sub> digested burgers showed the lowest polyphenol  
599 294 bioaccessibility (188 mg/100 g). All other digested samples did not show a statistically significant  
600 294 difference compared to digested CTRL burgers (214 mg/100 g).

601 295  
602 295 The antioxidant capacity of the burgers was measured before and after *in vitro* digestion using three  
603 296 different methods: ORAC (Fig 1B), ABTS (Fig 1C) and DPPH (Fig 1D). These methods were chosen  
604 297 in order to evaluate different aspects of the chemical mechanisms of action (Serpen, Gökmen, &  
605 297 Fogliano, 2012). In both cases (digested or non-digested), we obtained the following results: ORAC >  
606 298 ABTS > DPPH.  
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609 300 Before digestion, we found a good correspondence between antioxidant capacities and polyphenol  
610 300 patterns, especially for the ORAC and ABTS assays (Fig. 1A, B, C, blank bars). CTRL burgers showed  
611 301 interesting antioxidant capacity values, specifically 1104 (ORAC), 132 (ABTS) and 51 (DPPH)  
612 302  $\mu\text{mol}_{\text{TE}}/100 \text{ g}$ . The addition of goji puree and chia seeds in G<sub>5</sub>+C<sub>5</sub> burgers increased the antioxidant  
613 302 capacities of our burgers up to 1902 (ORAC), 236 (ABTS) and 132 (DPPH)  $\mu\text{mol}_{\text{TE}}/100 \text{ g}$ .

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615 303 After digestion, the antioxidant capacities of cooked burgers were all higher than non-digested samples,  
616 304 thus reflecting a higher polyphenol bioaccessibility (Fig. 1A, B, C, D grey bars). The ORAC and DPPH  
617 305 methods revealed a higher antioxidant capacity when goji puree and chia seeds were added,  
618 305 respectively, thus highlighting the different ability of polyphenols to scavenge free radicals.  
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### 621 307 622 308 **3.4. Lipid peroxidation in cooked beef burgers before and after the *in vitro* digestion**

623 309  
624 309 Unsaturated fatty acids in meat are susceptible to oxidation leading to a deterioration in quality, which  
625 310 may include color changes, off-flavors and odors. Several authors have demonstrated that lipid  
626 310 peroxidation in meat products could be prevented by the addition of protective compounds such as  
627 311 polyphenols (Gorelik, Ligumsky, Kohen, & Kanner, 2008), fibers (Hur, Lim, Park, & Joo, 2009),  
628 311 minerals and vitamins (Pierre et al., 2013).  
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631 313 We therefore investigated lipid peroxidation by measuring MDA formation on cooked burgers before  
632 313 and after *in vitro* digestion (Figure 2).  
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635 315 Before digestion (Fig. 2, blank bars), CTRL burgers showed the highest MDA value (0.66 mg<sub>MDA</sub>/100  
636 316 g). The addition of goji puree at 2.5% (G<sub>2.5</sub>) or 5% (G<sub>5</sub>) resulted in a gradual statistically significant  
637 316 MDA decrease (0.48 and 0.35 mg<sub>MDA</sub>/100 g, respectively), notwithstanding the possible interference of  
638 317 sugars and red pigments contained in this plant-based additive (Jung et al., 2016). The addition of chia  
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652 323 seeds also provided a significant reduction in MDA levels compared to CTRL burgers, though not in a  
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654 324 dose-dependent manner (0.30 mg<sub>MDA</sub>/100 g for both C<sub>2.5</sub> and C<sub>5</sub>). The protective effect against lipid  
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656 325 peroxidation is attributable to the high polyphenol and fiber content of goji puree (Kulczyński &  
657 326 Gramza-Michałowska, 2016; Zhou et al., 2017) and chia seeds (Alfredo, Gabriel, Luis, & David, 2009;  
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659 327 Ullah et al., 2016).  
660 328 After digestion (Fig. 2, grey bars), lipid peroxidation significantly increased in all samples (for  
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662 329 example, from 0.66 in non-digested to 2.13 mg<sub>MDA</sub>/100 g in digested CTRL burgers) because of the  
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664 330 oxidative processes occurring during digestion described in the literature (Kim & Hur, 2018; Martínez,  
665 331 Nieto, Castillo, & Ros, 2014). The addition of goji puree and chia seeds provided different results  
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667 332 regarding protection against lipid peroxidation, with the former showing good results when used at  
668 333 higher concentrations (G<sub>5</sub> = 1.46 mg<sub>MDA</sub>/100 g) and the latter when used at lower concentrations (C<sub>2.5</sub> =  
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670 334 1.57 mg<sub>MDA</sub>/100 g).  
671 335 It is well known that high-fat beef products are more susceptible to lipid peroxidation due to their  
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673 336 content of heme-Fe, which catalyzes the production of ROS and the oxidation of unsaturated fatty  
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675 337 acids. Beef burgers containing 5% chia seeds, used alone (C<sub>5</sub>) or in combination with goji puree  
676 338 (G<sub>5</sub>+C<sub>5</sub>), showed an important increase in MDA levels after *in vitro* digestion, thus suggesting a pro-  
677  
678 339 oxidative effect of the seeds. Indeed, chia seeds contain high levels of unsaturated fatty acids,  
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680 340 especially  $\alpha$ -linolenic acid (Muñoz et al., 2013), which is probably made more susceptible to lipid  
681 341 peroxidation by the *in vitro* digestion process. Moreover, the C<sub>5</sub> burger was the sample that showed the  
682  
683 342 lowest polyphenol bioaccessibility after digestion (Fig. 1A, grey bar), thus suggesting poor antioxidant  
684 343 protection against oxidation caused by the digestion process.

### 686 344 **3.5. Sensory acceptability**

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689 345 Considering the latest results regarding lipid peroxidation, we decided to perform the sensory analysis  
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691 346 on only four out of seven burgers (CTRL, G<sub>2.5</sub>, C<sub>2.5</sub>, G<sub>2.5</sub>+C<sub>2.5</sub>).

692 347 A significant effect of the subject group ( $P < 0.0001$ ) was found on liking for all the sensory  
693  
694 348 modalities. The elderly always showed significantly higher mean liking scores for all sensory  
695 349 modalities than adults or young people. The liking of the elderly was also significantly higher than  
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697 350 adults for odor, taste, texture and overall liking. Therefore, in general, liking decreased as follows:  
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699 351 elderly > adults > young. Acceptability (score = 5) was attained in all groups for all samples.

700 352 The mean liking values obtained for the four burgers by all assessors and the three related groups  
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702 353 (young, adult, elderly) are shown in Table 5.

703 354 Considering the whole population, samples only significantly affected liking as regards appearance,  
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705 355 with G<sub>2.5</sub> and CTRL showing the highest mean value.

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711 356 Within the groups, the effect of the sample on liking was different. In the young group, the sample had  
712 a significant ( $P < 0.01$ ) effect on liking as regards appearance, taste, flavor, texture and overall liking.  
713 357 Liking scores were generally modest in this group. For all sensory modalities, young people liked the  
714 G<sub>2.5</sub> sample at the same level as the control sample (CTRL). In adults, the effect of the sample was only  
715 358 observed on liking for flavor, with C<sub>2.5</sub> proving to be the most preferred sample. In the elderly, no  
716 359 significant effects of the sample on liking were observed for any of the sensory modalities. However,  
717 liking scores were generally higher than 6 (moderately liked); therefore, all samples showed a good  
718 360 performance regardless of the **formulation**.  
719 361 Results regarding the frequency of meat consumption (as reported in Materials and Methods section)  
720 showed the following distribution in the three classes: 13% occasional meat-eaters, 38% frequent meat-  
721 362 eaters, and 49% very frequent meat-eaters. The frequency of consumption of meat had a significant  
722 363 effect ( $P \leq 0.05$ ) on appearance, odor, taste, flavor, and texture but not on overall liking. In all cases,  
723 liking increased as the frequency of consumption increased: very frequent > frequent > occasional.  
724 364 Mean values of all sensory modalities were significantly higher for the very frequent consumers than  
725 for the occasional consumers. Significant different mean values between very frequent and frequent  
726 365 consumers were observed only in terms of liking as regards flavor.  
727 366 The high liking score results obtained from the hedonic tests were extremely encouraging. This is not  
728 always the case in functionalized meat products, as observed in beef patties formulated with flaxseeds,  
729 367 where an increase in the plant-based ingredient led to lower liking scores (Elif Bilek & Turhan, 2009).  
730 Moreover, the most positive results were found among the elderly, who could be considered as a  
731 368 particularly desirable consumer target for the developed burgers. In fact, elderly people often do  
732 369 not cover their protein needs. This situation is especially worrisome in retirement communities, where  
733 the majority of elderly residents do not meet their caloric and protein requirements and the institutions  
734 370 have to face the elderly's beliefs that 'At my age, I no longer need to eat so much meat' (Sulmont-  
735 371 Rossé, and Van Wymelbeke, 2019). The tests involving elderly subjects were conducted in this type of  
736 setting (retirement home); hence, these types of meats could have promising applications in the near  
737 372 future.  
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## 756 384 **Conclusions**

757  
758 385 The addition of goji berry puree and chia seeds affected the physical-chemical, nutritional and sensory  
759 386 properties of cooked beef burgers in different ways. Chia seeds make it possible to label burgers with  
760 EU health claims regarding fatty acids. Both plant-based ingredients ameliorated polyphenol content  
761 387 and total antioxidant capacity of beef burgers, before and after digestion, thus suggesting a better  
762 388 bioaccessibility of antioxidant molecules and a possible greater bioavailability. The addition of  
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770 390 antioxidants to the diet, especially if they are consumed at the same time as meat products or in the  
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772 391 same meat preparation, could be a good strategy to counteract the lipid peroxidation that usually occurs  
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774 392 in cooked meat products. Nevertheless, the concentration of the vegetables added to the burgers should  
775 393 be carefully dosed in order to avoid a pro-oxidative effect. Finally, sensory acceptability was attained  
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777 394 in all groups for all the samples, with the most interesting results obtained in the elderly target (<60  
778 395 years). This finding, together with all the improved nutritional qualities and antioxidant capacities,  
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780 396 suggest that these enhanced burgers could become a valid meal alternative for human nutrition.

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783 398 **Declarations of interest**

784  
785 399 None.

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787 400 **Acknowledgments**

788  
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793 404 **References**

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546 **Table 1.** Beef burger formulations.

<b>Ingredients (%)</b>	<b>CTRL</b>	<b>G<sub>2.5</sub></b>	<b>C<sub>2.5</sub></b>	<b>G<sub>5</sub></b>	<b>C<sub>5</sub></b>	<b>G<sub>2.5</sub>+C<sub>2.5</sub></b>	<b>G<sub>5</sub>+C<sub>5</sub></b>
Ground beef meat	85	85	85	85	85	85	85
Chia seeds (C)	0	0	2.5	0	5	2.5	5
Goji puree (G)	0	2.5	0	5	0	2.5	5
Vegetable dietary fibre	2	2	2	2	2	2	2
Salt	0.5	0.5	0.5	0.5	0.5	0.5	0.5
Water	12.5	10	10	7.5	7.5	7.5	2.5
<b>Total</b>	<b>100</b>	<b>100</b>	<b>100</b>	<b>100</b>	<b>100</b>	<b>100</b>	<b>100</b>

**Table 2.** Proximate composition, pH and cooking loss of beef burgers.

	Samples*						
	CTRL	G <sub>2.5</sub>	C <sub>2.5</sub>	G <sub>5</sub>	C <sub>5</sub>	G <sub>2.5</sub> +C <sub>2.5</sub>	G <sub>5</sub> +C <sub>5</sub>
<b>Proximate analysis</b>							
Moisture (%)	64.7 ± 0.5 <sup>a</sup>	64.3 ± 0.3 <sup>a</sup>	64.1 ± 0.1 <sup>a</sup>	63.0 ± 0.5 <sup>a</sup>	61.9 ± 0.3 <sup>b</sup>	63.1 ± 0.5 <sup>a</sup>	60.7 ± 0.5 <sup>b</sup>
Fat (%)	12.3 ± 0.1 <sup>c</sup>	12.0 ± 0.1 <sup>c</sup>	13.8 ± 0.1 <sup>b</sup>	11.8 ± 0.1 <sup>c</sup>	14.7 ± 0.2 <sup>a</sup>	13.8 ± 0.1 <sup>b</sup>	14.0 ± 0.2 <sup>b</sup>
Protein (%)	23.5 ± 0.5 <sup>a</sup>	23.2 ± 0.3 <sup>a</sup>	22.7 ± 0.2 <sup>a</sup>	23.3 ± 0.3 <sup>a</sup>	22.4 ± 0.2 <sup>a</sup>	22.3 ± 0.3 <sup>a</sup>	22.0 ± 0.4 <sup>a</sup>
pH	5.9 ± 0.0 <sup>a</sup>	5.8 ± 0.0 <sup>b</sup>	5.8 ± 0.0 <sup>b</sup>	5.7 ± 0.0 <sup>c</sup>	5.8 ± 0.0 <sup>b</sup>	5.8 ± 0.0 <sup>b</sup>	5.7 ± 0.0 <sup>c</sup>
CL (%)	26.1 ± 1.2 <sup>a</sup>	24.2 ± 1.0 <sup>a</sup>	21.2 ± 0.6 <sup>b</sup>	24.5 ± 0.8 <sup>a</sup>	16.7 ± 0.8 <sup>c</sup>	19.8 ± 0.8 <sup>b</sup>	18.7 ± 0.3 <sup>b</sup>

\*For sample formulations see Table 1. CL, cooking loss. Values are reported as mean ± standard error.

<sup>a,b</sup>Different letters in the same row indicate statistically significant differences ( $P < 0.05$ ).

**Table 3.** Fatty acid profiles and nutritional significance ratios on beef burgers.

Parameters	Samples*						
	CTRL	G <sub>2.5</sub>	C <sub>2.5</sub>	G <sub>5</sub>	C <sub>5</sub>	G <sub>2.5</sub> +C <sub>2.5</sub>	G <sub>5</sub> +C <sub>5</sub>
<b>C14:0</b>	3.1 ± 0.1	3.2 ± 0.1	3.1 ± 0.1	3.2 ± 0.1	2.9 ± 0.1	3.1 ± 0.1	2.9 ± 0.1
<b>C 16:0</b>	27.3 ± 0.5	26.8 ± 0.5	25.5 ± 0.5	26.8 ± 0.5	25.0 ± 0.5	25.6 ± 0.5	24.4 ± 0.5
<b>C 18:0</b>	15.1 ± 0.4	15.9 ± 0.4	15.0 ± 0.4	15.8 ± 0.4	14.7 ± 0.4	15.0 ± 0.4	14.0 ± 0.4
<b>Total SFA</b>	<b>45.5 ± 0.7<sup>a</sup></b>	<b>45.9 ± 0.7<sup>a</sup></b>	<b>43.5 ± 0.7<sup>b</sup></b>	<b>45.8 ± 0.7<sup>a</sup></b>	<b>42.6 ± 0.7<sup>b</sup></b>	<b>43.7 ± 0.7<sup>b</sup></b>	<b>41.2 ± 0.7<sup>b</sup></b>
<b>C14:1</b>	1.1 ± 0.0	1.0 ± 0.0	1.0 ± 0.0	1.0 ± 0.0	0.9 ± 0.0	1.0 ± 0.0	1.0 ± 0.0
<b>C16:1</b>	4.0 ± 0.2	3.9 ± 0.2	3.8 ± 0.2	3.9 ± 0.2	3.5 ± 0.1	3.8 ± 0.2	3.6 ± 0.2
<b>C18:1</b>	47.9 ± 0.8	46.7 ± 0.8	45.0 ± 0.7	46.7 ± 0.8	43.0 ± 0.7	45.0 ± 0.7	43.5 ± 0.7
<b>Total MUFA</b>	<b>52.9 ± 0.8<sup>a</sup></b>	<b>51.6 ± 0.8<sup>a</sup></b>	<b>49.8 ± 0.8<sup>ab</sup></b>	<b>51.6 ± 0.8<sup>a</sup></b>	<b>47.5 ± 0.7<sup>b</sup></b>	<b>49.8 ± 0.8<sup>ab</sup></b>	<b>48.1 ± 0.7<sup>b</sup></b>
<b>C18:2 (ω-6)</b>	2.4 ± 0.1	2.4 ± 0.1	3.4 ± 0.1	2.4 ± 0.1	4.1 ± 0.2	3.3 ± 0.1	4.3 ± 0.2
<b>C18:3 (ω-3)</b>	0.4 ± 0.0	0.4 ± 0.0	3.6 ± 0.2	0.4 ± 0.0	6.1 ± 0.2	3.4 ± 0.1	6.6 ± 0.3
<b>Total PUFA</b>	<b>2.9 ± 0.1<sup>c</sup></b>	<b>2.8 ± 0.1<sup>c</sup></b>	<b>7.0 ± 0.2<sup>b</sup></b>	<b>2.8 ± 0.1<sup>c</sup></b>	<b>10.1 ± 0.3<sup>a</sup></b>	<b>6.7 ± 0.2<sup>b</sup></b>	<b>10.9 ± 0.3<sup>a</sup></b>
PUFA/SFA	0.06	0.06	0.16	0.06	0.24	0.15	0.26
ω-6/ω-3	5.67	6.34	0.93	5.90	0.67	0.96	0.65
AI	0.60	0.61	0.56	0.61	0.53	0.56	0.51
TI	1.56	1.62	1.13	1.61	0.94	1.15	0.87

\*For sample formulations see Table 1. SFA, saturated fatty acids; MUFA, monounsaturated fatty acids; PUFA, polyunsaturated fatty acids; AI, atherogenicity index; TI, thrombogenicity index. Fatty acid profiles (percentages of total fat) are reported as average values ± standard error. <sup>a,b</sup>Different letters in the same row indicate statistically significant differences ( $P < 0.05$ ).

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**Table 4.** Antioxidant profiles of goji puree and chia seeds.

	Goji puree	Chia seeds
TPs (mg <sub>GAE</sub> /100 g)	153 ± 5 <sup>a</sup>	151 ± 2 <sup>a</sup>
ORAC (μmol <sub>TE</sub> /100 g)	9950 ± 29 <sup>a</sup>	6500 ± 23 <sup>b</sup>
ABTS (μmol <sub>TE</sub> /100 g)	2059 ± 101 <sup>a</sup>	1227 ± 55 <sup>b</sup>
DPPH (μmol <sub>TE</sub> /100 g)	213 ± 13 <sup>b</sup>	659 ± 8 <sup>a</sup>

TPs, total phenols. Values are reported as mean ± standard error. <sup>a,b</sup>Different letters in the same row indicate statistically significant differences ( $P < 0.05$ ).

**Table 5.** Overall liking and liking for appearance, odor, taste, flavor and texture obtained by the totality of the subjects and the three groups of consumers (young, adults, elderly) for burger samples.

Group	Samples*	Appearance	Odor	Taste	Flavor	Texture	Overall
All (n=127)	CTRL	6.0 ± 0.1 <sup>a</sup>	5.6 ± 0.1 <sup>a</sup>	5.9 ± 0.1 <sup>a</sup>	5.8 ± 0.1 <sup>ab</sup>	6.2 ± 0.1 <sup>a</sup>	6.0 ± 0.1 <sup>ab</sup>
	G <sub>2.5</sub> +C <sub>2.5</sub>	5.6 ± 0.1 <sup>b</sup>	5.4 ± 0.1 <sup>a</sup>	5.7 ± 0.1 <sup>a</sup>	5.6 ± 0.1 <sup>b</sup>	5.9 ± 0.1 <sup>a</sup>	5.8 ± 0.1 <sup>b</sup>
	G <sub>2.5</sub>	6.0 ± 0.1 <sup>a</sup>	5.7 ± 0.1 <sup>a</sup>	6.1 ± 0.1 <sup>a</sup>	6.0 ± 0.1 <sup>a</sup>	6.2 ± 0.1 <sup>a</sup>	6.1 ± 0.1 <sup>a</sup>
	C <sub>2.5</sub>	5.5 ± 0.1 <sup>b</sup>	5.6 ± 0.1 <sup>a</sup>	5.8 ± 0.1 <sup>a</sup>	5.8 ± 0.1 <sup>ab</sup>	6.0 ± 0.1 <sup>a</sup>	5.9 ± 0.1 <sup>ab</sup>
Young (n=56)	CTRL	5.8 ± 0.1 <sup>a</sup>	5.3 ± 0.1 <sup>a</sup>	6.0 ± 0.1 <sup>a</sup>	5.7 ± 0.1 <sup>ab</sup>	5.9 ± 0.2 <sup>a</sup>	5.8 ± 0.1 <sup>a</sup>
	G <sub>2.5</sub> +C <sub>2.5</sub>	4.6 ± 0.1 <sup>b</sup>	4.8 ± 0.1 <sup>b</sup>	5.3 ± 0.1 <sup>b</sup>	5.1 ± 0.1 <sup>c</sup>	5.2 ± 0.2 <sup>bc</sup>	5.1 ± 0.1 <sup>c</sup>
	G <sub>2.5</sub>	5.8 ± 0.1 <sup>a</sup>	5.3 ± 0.1 <sup>a</sup>	6.0 ± 0.1 <sup>a</sup>	5.8 ± 0.1 <sup>a</sup>	5.7 ± 0.2 <sup>ab</sup>	5.8 ± 0.1 <sup>a</sup>
	C <sub>2.5</sub>	4.5 ± 0.1 <sup>b</sup>	4.9 ± 0.1 <sup>ab</sup>	5.1 ± 0.1 <sup>b</sup>	5.1 ± 0.1 <sup>bc</sup>	5.0 ± 0.2 <sup>c</sup>	5.1 ± 0.1 <sup>b</sup>
Adult (n=38)	CTRL	6.1 ± 0.1 <sup>a</sup>	5.6 ± 0.1 <sup>a</sup>	5.6 ± 0.1 <sup>b</sup>	5.7 ± 0.1 <sup>b</sup>	6.2 ± 0.1 <sup>ab</sup>	5.9 ± 0.1 <sup>b</sup>
	G <sub>2.5</sub> +C <sub>2.5</sub>	6.2 ± 0.1 <sup>a</sup>	5.5 ± 0.1 <sup>a</sup>	5.8 ± 0.1 <sup>ab</sup>	5.9 ± 0.1 <sup>b</sup>	6.1 ± 0.1 <sup>b</sup>	6.0 ± 0.1 <sup>ab</sup>
	G <sub>2.5</sub>	5.8 ± 0.1 <sup>a</sup>	5.5 ± 0.1 <sup>a</sup>	5.6 ± 0.1 <sup>b</sup>	5.8 ± 0.1 <sup>b</sup>	6.2 ± 0.1 <sup>ab</sup>	6.0 ± 0.1 <sup>ab</sup>
	C <sub>2.5</sub>	6.2 ± 0.1 <sup>a</sup>	5.8 ± 0.1 <sup>a</sup>	6.2 ± 0.1 <sup>a</sup>	6.4 ± 0.1 <sup>a</sup>	6.6 ± 0.1 <sup>a</sup>	6.4 ± 0.1 <sup>a</sup>
Elderly (n=33)	CTRL	6.1 ± 0.1 <sup>b</sup>	6.1 ± 0.2 <sup>b</sup>	6.2 ± 0.2 <sup>a</sup>	6.2 ± 0.2 <sup>a</sup>	6.8 ± 0.1 <sup>a</sup>	6.3 ± 0.1 <sup>a</sup>
	G <sub>2.5</sub> +C <sub>2.5</sub>	6.6 ± 0.1 <sup>a</sup>	6.4 ± 0.2 <sup>a</sup>	6.5 ± 0.2 <sup>a</sup>	6.2 ± 0.2 <sup>a</sup>	6.8 ± 0.1 <sup>a</sup>	6.7 ± 0.1 <sup>a</sup>
	G <sub>2.5</sub>	6.6 ± 0.1 <sup>a</sup>	6.6 ± 0.2 <sup>a</sup>	6.8 ± 0.2 <sup>a</sup>	6.8 ± 0.2 <sup>a</sup>	7.0 ± 0.1 <sup>a</sup>	6.8 ± 0.1 <sup>a</sup>
	C <sub>2.5</sub>	6.4 ± 0.1 <sup>ab</sup>	6.4 ± 0.2 <sup>ab</sup>	6.5 ± 0.2 <sup>a</sup>	6.1 ± 0.2 <sup>a</sup>	6.8 ± 0.1 <sup>a</sup>	6.7 ± 0.1 <sup>a</sup>

\*For sample formulations see Table 1. Values are reported as mean ± standard error. Different letters among the same sub-column indicate statistically significant differences ( $P < 0.05$ ).

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569**Table S1.** Average nutritional values of goji puree and chia seeds used in beef burger formulations.

	Goji puree	Chia seeds
<b>Nutritional values*</b>		
Energy (kJ and kcal/100 g)	337 / 80	1992 / 482
Fat (g/100 g)	0.91	35
of which saturates (g/100 g)	0.43	3.5
Carbohydrate (g/100 g)	13	5.3
of which sugars (g/100 g)	9.5	0.55
Dietary fiber (g/100 g)	5.4	29
Protein (g/100 g)	2.2	21
Salt (g/100 g)	0.13	0.05
<b>Moisture (%)</b>	<b>76.8 ± 0.9<sup>a</sup></b>	<b>5.5 ± 0.4<sup>b</sup></b>
<b>pH</b>	<b>4.8 ± 0.0<sup>b</sup></b>	<b>7.8 ± 0.0<sup>a</sup></b>

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\*Average nutritional values as reported by the manufacturers on the label. Values of moisture and pH are reported as mean ± **standard error**. <sup>a,b</sup>Different letters in the same row indicate statistically significant differences ( $P < 0.05$ ).

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**Table S2.** Three-stage *in vitro* digestion procedure (A) and composition of digestive juices (B).

<b>A) Digestion phases</b>				
	Sample	Extraction mixture	Time (min)	Temperature (°C)
Oral	Cooked beef burgers (5 g)	6 ml of salivary solution (pH = 6.8)	5	37
Gastric	Suspension after the oral phase	12 ml of gastric solution (pH = 1.5)	120	37
Intestinal	Suspension after the gastric phase	12 ml of duodenal + 6 ml of biliar + 1 M bicarbonate solutions (pH = 8.2)	120	37

<b>B) Constituents and composition of 1L of salivary, gastric, duodenal and biliar juices</b>				
	Salivary	Gastric	Duodenal	Biliar
<b>Organic and inorganic components</b>	1.7 ml NaCl (175.3 g/L)	6.5 ml HCl (37 %)	6.3 ml KCl (89.6 g/L)	68.5 ml NaHCO <sub>3</sub> (84.7 g/L)
	8.0 ml urea (25.0 g/L)	18.0 ml CaCl <sub>2</sub> (22.2 g/L)	9.0 ml CaCl <sub>2</sub> (22.2 g/L)	10.0 ml CaCl <sub>2</sub> (22.2 g/L)
	15 mg uric acid			
<b>Enzymes</b>	290 mg $\alpha$ -amilasi 25 mg mucin	2.5 g pepsin 3 g mucin 1 g BSA	1.5 g lipase 9 g pancreatin 1 g BSA	30 g bile 1.8 g BSA

After mixing all ingredients (organic and inorganic components and enzymes), the volume was increased to 1 L with distilled water. If necessary, the pH of the juices was adjusted to the appropriate value.



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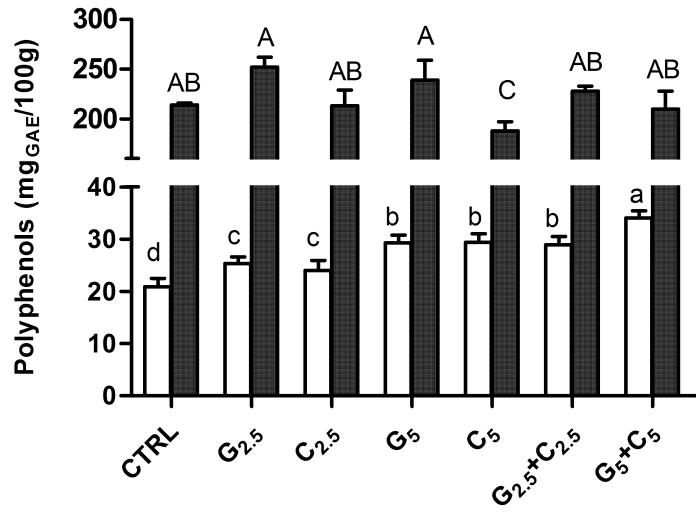
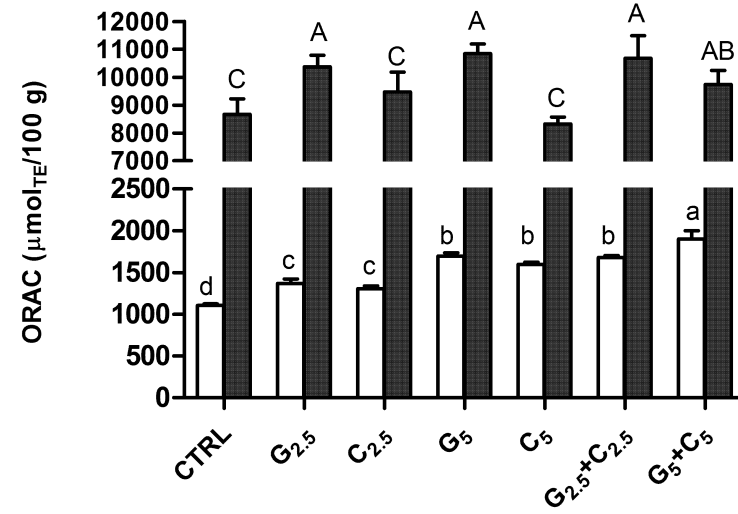
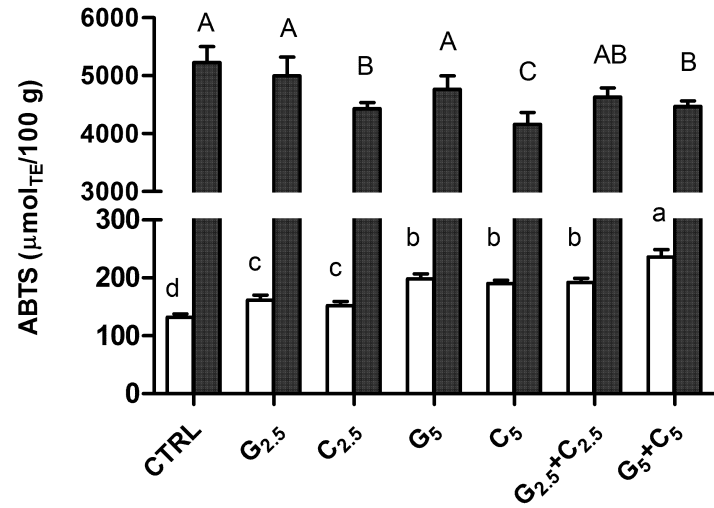
**Figure captions**

**Figure 1.** Polyphenols (A), ORAC (B), ABTS (C), DPPH (D) of cooked beef burgers before (blank bars) and after (grey bars) *in vitro* digestion process.

For sample formulations see Table 1. Values are reported as mean  $\pm$  standard error. Different letters indicate statistically significant differences among non-digested (a, b) or digested (A, B) samples ( $P < 0.05$ ).

**Figure 2.** Lipid peroxidation of cooked beef burgers before (blank bars) and after (grey bars) *in vitro* digestion process.

For sample formulations see Table 1. Values are reported as mean  $\pm$  standard error. Different letters indicate statistically significant differences among non-digested (a, b) or digested (A, B) samples ( $P < 0.05$ ).

**A****B****C****D**